

### Seshat output (long)

<b>cDNA_Variant</b>	Mutation nomenclature according to HGVS standards using the coding sequence as reference (position 1 refers to the A of the start ATG): reference sequence <a href="#">NM_000546.5</a>
<b>HG19_Variant</b>	Mutation nomenclature according to HGVS standards using the genomic sequence as reference Reference sequence: NC_000017.10 for genome build NCBI37/hg19
<b>NG_017013.2_variant</b>	Mutation nomenclature according to HGVS standards using the RefSeq Gene NG_017013. sequence as reference <a href="http://www.ncbi.nlm.nih.gov/nuccore/NG_017013.2">http://www.ncbi.nlm.nih.gov/nuccore/NG_017013.2</a> This sequence is also the reference used by the Locus Reference Genomic ( <a href="http://ftp.ebi.ac.uk/pub/databases/lrgex/LRG_321.xml">http://ftp.ebi.ac.uk/pub/databases/lrgex/LRG_321.xml</a> )
<b>SNP_ID*</b>	<p>The SNP database now includes several pathogenic variants of the TP53 gene <a href="http://www.ncbi.nlm.nih.gov/snp">http://www.ncbi.nlm.nih.gov/snp</a></p> <p>* <b>Note of caution:</b> Since 2011 (build 134), dbSNP started accepting submissions of germ line and somatic variations associated with various types of diseases and changed its name to “database of Short Genetic Variation” keeping the dbSNP acronym. Several frequent <i>TP53</i> variants (rs121912651, c.742C&gt;T, p.Arg248Trp or rs11540652, c.743G&gt;A, p.Arg248Gln) are included in dbSNP, but other hot spot variants are missing, whereas rare somatic variants <b>may be included</b>. This heterogeneity caused by biased dbSNP submissions is misleading, as it does not reflect the true occurrence and frequencies of <i>TP53</i> variants. Therefore, without further distinction, we can no longer assume that variants in dbSNP are associated with the lack of effect on disease and tumour characteristics</p> <p>Common SNPs such as rs1042522 (p.P72R), rs1800371 (p.P47S), rs1800372 (p.R213R) or rs1800370 (p.P36P) are not included in the database..</p>
<b>Transcript t1 MN_000546.5</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference sequence NM_000546.5</i>
<b>Transcript t2 NM_001126112.2</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference sequence NM_001126112.2</i>
<b>Transcript t3 NM_001126114.2</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference sequence NM_001126114.2</i>
<b>Transcript t4 NM_001126113.2</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference sequence NM_001126113.2</i>
<b>Transcript t5 NM_001126115.1</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference sequence NM_001126115.1</i>
<b>Transcript t6 NM_001126116.1</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference sequence NM_001126116.1</i>
<b>Transcript t7 NM_001126117.1</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference sequence NM_001126117.1</i>
<b>Transcript t8 NM_001126118.1</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG (position 1 refers to the A of the start ATG): reference</i>

	<i>sequence NM_001126118.1</i>
<b>Protein p1 TP53_alpha</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p1</i>
<b>Protein p3 TP53_beta</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p3</i>
<b>Protein p4 TP53_gamma</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p4</i>
<b>Protein p8 Delta40_TP53_alpha</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p8</i>
<b>Protein p9 Delta 40_TP53_beta</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p9</i>
<b>Protein p10 Delta 40_TP53_gamma</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p10</i>
<b>Protein p5 Delta 133_TP53_alpha</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p5</i>
<b>Protein p6 Delta 133_TP53_beta</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence LRG_321p6</i>
<b>Protein p7 Delta 133_TP53_gamma</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence NP_001119589.1</i>
<b>Protein p11 Delta160_TP53_alpha</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence NP_001263626.1</i>
<b>Protein p12 Delta160_TP53_beta</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence NP_001263627.1</i>
<b>Protein p13 Delta160_TP53_gamma</b>	<i>Mutation nomenclature and coordinates are described according to HGVS, NCBI and LRG: reference sequence NP_001263628.1</i>
<b>Records_Number</b>	Number of occurrences of the mutant in the database.
<b>Variant_Classification</b>	<p>Translational effect of the mutation (Missense, Nonsense, Synonymous, Nonstop, In_frame_Del, Inframe_Ins, Frameshift_Del or Frameshift_Ins).</p> <p>These 8 items are identical to the entries used in MAF file</p> <p>Four novel items are used in the TP53 mutation database: In_frame_Del_Complex, Inframe_Ins_Complex, Frameshift_Del_Complex or Frameshift_Ins_Complex) for mutations that span one or more than one exon-intron site.</p>
<b>Variant_Type</b>	<p>Variant type as defined in MAF file</p> <p><b>SNV:</b> Single Nucleotide Variant  <b>DNP:</b> Change in two consecutive bases (dinucleotide variant)  <b>TNP:</b> Change in three consecutive bases (tri-nucleotide variant)  <b>ONP:</b> Change in four or more consecutive bases (oligo-nucleotide variant)  <b>INS:</b> Insertion  <b>DEL:</b> Deletion</p>
<b>Comment_1_Frequency</b>	<p>Specific information related to the frequency of the mutation in the database.</p> <p>Four categories have been defined:</p> <p>i: This mutation is very frequent  ii) This mutation is frequent  iii: This mutation is not frequent  iv: This mutation is rare</p> <p>see Leroy et al. TP53 Mutations in Human Cancer: Database Reassessment and Prospects for the Next Decade. Human Mutation (2014) 35, 672-688</p>

<b>Comment_2_Activity</b>	<p>Specific information related to the residual activity of this TP53 mutant in the database based on the overall transcriptional activity (TA) on 8 different promoters as measured by Kato et al. For each mutant, the median of the 8 promoter-specific activities (expressed as percent of the wild-type protein) has been calculated.</p> <p>For <b>missense variants</b>, five categories have been defined:</p> <ul style="list-style-type: none"> <li>• No activity: median <math>\leq 20</math></li> <li>• Partial activity: median <math>&gt; 20</math> and <math>\leq 75</math></li> <li>• Fully active: median <math>&gt; 75</math> and <math>\leq 140</math></li> <li>• Hyper active: median <math>&gt; 140</math></li> <li>• No data: this mutant has not been tested</li> </ul> <p>For <b>nonsense variants</b>, one category has been used The activity of truncated p53 is assumed to be nil</p> <p>For <b>frameshift</b> variants, two categories have been used:</p> <p>The consequence of this in-frame mutation is unknown (In-frame of 15 bp or less). The activity of truncated p53 is assumed to be nil (out-of-frame insertion and deletion, in-frame mutation <math>&gt; 18</math> bp or mutation across an intron:exon junction).</p> <p>For <b>synonymous</b> variants, two categories have been used:</p> <p>This synonymous mutation is known to impair TP53 splicing. Synonymous mutation with unknown consequences.</p> <p>For mutations that target the canonical AG <b>splice-acceptor site</b> or GT <b>splice-donor site</b>:</p> <p>Splicing defect: impaired TP53 activity</p> <p>Activity for each individual promoter is also available (see the various rows in the database: WAF, MDM2, BAX, 14-3-3-s, AIP, GADD45, NOXA and P53R2).</p>
<b>Comment_3_Isoforms</b>	Number of TP53 isoforms targeted by the mutation
<b>Comment_4_Prediction</b>	<p>Several prediction algorithms have been used to predict TP53 loss of function (SIFT, Mutassessor, Provean, PolyPhen, see the corresponding rows for each individual analysis).</p> <p>A prediction index has been deduced from the various analyses</p> <p>Damaging Probably damaging Tolerated</p> <p>Note of caution: for TP53 mutation, the sensitivity of the various algorithms is never higher than 80% and the relation between loss of function and pathogenicity is not straightforward.</p>
<b>Comment_5_Outliers</b>	<p>Indicates whether or not the mutation is associated with outlier publications. See Edlund K, Larsson O, Ameur A, Bunikis I, Gyllenstein U, Leroy B, Sundstrom M, Micke P, Botling J, Soussi T. 2012. Data-driven unbiased curation of the TP53 tumor suppressor gene mutation database and validation by ultradeep sequencing of human tumors. Proc Natl Acad Sci USA 109:9551–9556. for more info on outliers studies.</p>

	Rare mutants only found in outlier studies should be considered to be suspicious.
<b>Comment_6_Splicing</b>	<p>Indicates whether or not the mutation could impair TP53 splicing.</p> <p>All TP53 gene substitutions have been analysed by using mutpred_splice M. Mort <i>et al.</i>, <i>Genome Biol</i> <b>15</b>, R19 (2014) courtesy of M. Mort</p> <p>A MutPred Splice general score probability cutoff of <math>\geq 0.70</math> was used to indicate a predicted SAV.</p> <p>For mutations close to an exon, a cutoff of <math>\geq 0.60</math> was used.</p> <p>Raw data for mutpred_splice are also available in this table.</p>
<b>Comment_7_Sequence</b>	Indicates the presence of homopolymeric tracts at the position of the mutation.
<b>Comment_9_SNP</b>	Specific comments regarding the specificity of each SNP including novel SNP detected in new sequencing projects
<b>Comment_10_population</b>	Population data and frequency of the SNP in various databases.
<b>Pathogenicity</b>	<p>We used this specific standard terminology for TP53 variants: 'pathogenic', 'likely pathogenic', 'uncertain significance' (VUS), 'likely benign'.</p> <p>A new TP53 specific algorithm was used to define TP53 variant pathogenicity (T Soussi et al. manuscript in preparation).</p>
<b>Final comment</b>	Comment summary.
<b>HG18_Variant</b>	<p>Mutation nomenclature according to HGVS standards using the genomic sequence as reference</p> <p>Reference sequence: NC_000017.9 for genome build NCBI36/hg18</p>
<b>HG38_Variant</b>	<p>Mutation nomenclature according to HGVS standards using the genomic sequence as reference</p> <p>Reference sequence: NC_000017.11 for genome build GRCh38.p2</p>
<b>UMD_ID</b>	Unique mutation identifier used in the UMD database for each genomic variant
<b>COSMIC_ID</b>	<p>Mutation identifier used in COSMIC</p> <p><a href="http://cancer.sanger.ac.uk/cancergenome/projects/cosmic/">http://cancer.sanger.ac.uk/cancergenome/projects/cosmic/</a></p>
<b>HG19 Start</b>	Mutation start coordinates using HG19 as reference
<b>HG19 End</b>	Mutation end coordinates using HG19 as reference
<b>HG18 Start</b>	Mutation start coordinates using HG18 as reference
<b>HG18 End</b>	Mutation end coordinates using HG18 as reference
<b>Exon:intron_Start</b>	<p>Location of the mutation start in the introns or exons of the TP53 gene. In most cases, Exon:intron_Start and Exon:intron_stop are similar.</p> <p>A few large deletions encompass several exons and introns.</p> <p>Although all intronic variations described in the literature have been included in the database, only mutations that target the canonical AG splice-acceptor site or GT splice-donor site (-1,-2, +1, +2) are considered to be pathogenic (see splice comment for more information).</p>
<b>Exon:intron_End</b>	<p>Location of the mutation end in the introns or exons of the TP53 gene. In most cases, Exon:intron_Start and Exon:intron_stop are similar.</p> <p>A few large deletions encompass several exons and introns.</p> <p>Although all intronic variations described in the literature have been included in the database, only mutations that target the canonical AG splice-acceptor site or GT splice-donor site (-1,-2, +1, +2) are considered to be pathogenic (see splice comment for more information).</p>
<b>Start_cDNA</b>	Mutation start coordinate using the p53 cDNA as reference (position 1 refers to the A of the start ATG): reference sequence <a href="#">NM_000546.5</a>
<b>End_cDNA</b>	Mutation end coordinate using the p53 cDNA as reference (position 1 refers to the A of the start ATG): reference sequence <a href="#">NM_000546.5</a>
<b>Genome base coding</b>	Nucleotide at the start position of the mutation.
<b>Mutant_Allele</b>	Mutant nucleotide

	<p>For deletion, this field is empty</p> <p>For insertion of unknown nucleotides, this field is empty</p> <p>For insertion, this field includes the inserted sequence except when this sequence is unknown and is therefore left empty.</p>
<b>Base_Change_Size</b>	Size of the substitution
<b>Ins_Size</b>	Size of the deletion
<b>Del_Size</b>	Size of the insertion; the sequence of the insertion is available for a few cases
<b>Codon</b>	<p><b>1-393:</b> Codon position using TP53 alpha (p1) as reference (NP_000537.2)</p> <p><b>Splice:</b> mutations that target the canonical AG acceptor site or GT donor.</p> <p><b>Untranslated:</b> mutations that target other nucleotides (5'UTR; 3'UTR or Intron)</p> <p>Large deletions with unknown boundaries are shown as "?".</p> <p>*: stop codon</p> <p>nnn-beta or nnn-gamma: Codon position specific for isoforms beta and gamma</p>
<b>WT_Codon</b>	<p>Nucleotide sequence of the wild-type codon in which the mutation occurred.</p> <p>Mutations that target the canonical AG splice-acceptor site or GT splice-donor site are displayed as <b>intron_nn_SA</b> or <b>intron_nn_SA</b>, where nn is the intron number</p> <p>For intronic mutations, the intron number is displayed (<b>intron_01 to intron 10, intron_09_beta and intron_09_gamma</b>)</p> <p>Large deletions with unknown boundaries are shown as “?”</p>
<b>Mutant_Codon</b>	<p><b>NNN:</b> Sequence of the mutated codon.</p> <p><b>Del:</b> exonic deletion</p> <p><b>Ins:</b> exonic insertion</p> <p><b>Indel:</b> complex event that involves an exonic insertion and a deletion. In accordance with the new HGVS rules, all <u>tandem mutations</u> are now included in this category as del2ins2 events. See the HGVS website for more information (<a href="http://www.hgvs.org/mutnomen/">http://www.hgvs.org/mutnomen/</a>).</p> <p><b>Splice:</b> mutation that targets the canonical AG splice-acceptor site or GT splice-donor site.</p> <p><b>NR:</b> not relevant, mutations targeting intronic sequence, 5'UTR or 3'UTR.</p>
<b>WT_AA_1</b>	<p>Wild-type amino acid: 1-letter nomenclature.</p> <p>For intronic mutations, the intron number is displayed (<b>intron_01 to intron 10, intron_09_beta and intron_09_gamma</b>).</p> <p>Mutations that target the canonical AG splice-acceptor site or GT splice-donor site are displayed as <b>intron_nn_SA</b> or <b>intron_nn_SA</b>, where nn is the intron number.</p>
<b>WT_AA_3</b>	<p>Wild-type amino acid: 3-letter nomenclature.</p> <p>For intronic mutations, the intron number is displayed (<b>intron_01 to intron 10, intron_09_beta and intron_09_gamma</b>)</p> <p>Mutations that target the canonical AG splice-acceptor site or GT splice-donor site are displayed as <b>intron_nn_SA</b> or <b>intron_nn_SA</b>, where nn is the intron number.</p>
<b>Mutant_AA_1</b>	Mutant amino acid: 1-letter nomenclature
<b>Mutant_AA_3</b>	Mutant amino acid: 3-letter nomenclature
<b>Substitution_Type</b>	<p><b>Ts:</b> Transition (a pyrimidine (C or T) is substituted by another pyrimidine, or a purine (A or G) is substituted by another purine);</p> <p><b>Tv:</b> Transversion (a transversion mutation involves substitution of a</p>

	pyrimidine by a purine, or vice versa); <b>Td</b> : tandem mutation <b>Fr</b> : Frameshift mutations (deletions / insertions) <b>Inf</b> : In-frame deletions or insertions
<b>CpG</b>	<b>Yes</b> : transition (G to A or C to T base change) at a CpG dinucleotide; <b>No</b> : transitions (G to A or C to T base change) at non-CpG sites and all transversions.
<b>Mutational_Event</b>	Mutational events <b>G&gt;C</b> : (G to C base substitution) All other single substitutions are described in a similar way. <b>CC&gt;TT</b> : mutation that changes two contiguous nucleotides. <b>Insertion</b> <b>Deletion</b> <b>Indel</b> : complex event that involves an insertion and a deletion.  <b>Important note: only the coding strand of the TP53 gene is used for mutation description.</b>
<b>Tandem_Class</b>	The majority of tandem mutations are found in skin tumours. Several types of tandem mutations can occur in the open reading frame of the TP53 gene (or any other genes). These mutations are considered to be single mutational events linked to UV exposure  <b>T1</b> : two different codons are modified by the substitution. e.g.: codons 247 and 248 of the TP53 gene: AAC - CCG -> AAT- AGG c.741_742delCCInsTA (p.[N247N; R248R]) In the majority of T1 tandem mutations, the first substitution does not change the amino acid residue and results in a synonymous change.  <b>T2</b> : only one codon is modified by the substitution. e.g.: codon 331, CAG ->CCA. c.992_993delAGInsCA (p.Q331P).  <b>T3</b> : Intronic tandem mutation that occurs across a splice site (+1/+2 or -1/-2)
<b>Mutation type</b>	<b>SNV</b> : Single nucleotide variant <b>D</b> : Deletion <b>I</b> : insertion <b>ID</b> : complex event that involves an insertion and a deletion.
<b>Variant_comment</b>	Specific comment concerning the consequences of the mutation.
<b>Domain</b>	Domain of the TP53 protein <ul style="list-style-type: none"> <li>• HCD I to V: Highly Conserved Domain I to V</li> <li>• DNA Binding: DNA binding domain</li> <li>• Negative regulation: carboxy-terminus of the p53 protein associated with negative regulation of p53 DNA binding activity</li> <li>• Transactivation TAD1: transactivation domain 1</li> <li>• Transactivation TAD2: transactivation domain 2</li> <li>• Proline Rich: Proline-rich domain of the p53 protein</li> <li>• NES: Nuclear export signal of p53</li> <li>• NLS: Nuclear localization signal of p53</li> <li>• Oligomerization: Tetramerization domain of the p53 protein</li> <li>• Empty field: No specific domain available</li> </ul>
<b>Structure</b>	Structural motif of the TP53 protein according to the analysis described by <a href="#">Cho et al. (1994)</a> .

<b>PTM</b>	Post-translational modifications <ul style="list-style-type: none"> <li>• Lys Acetylation</li> <li>• Lys Ubiquitination</li> <li>• Asp and Glu ADP Ribosylation</li> <li>• Ser or Thr Phosphorylation</li> <li>• Ser O-Linked Glycosylation</li> <li>• Cys Glutathionylation</li> <li>• Arg and Lys Methylation</li> <li>• Asn Isoaspartyl methylation</li> <li>• Lys Neddylation</li> <li>• Tyr Nitrosylation</li> <li>• Lys Methylation</li> </ul>
<b>Leukaemia_Lymphoma_Freq</b>	Frequency of the variant (cDNA_nomenclature) in haematological malignancies  e.g.: for variant c.524G>A, the entry will be 2.91 (216/7,403) <ul style="list-style-type: none"> <li>• Frequency of c.524G&gt;A in haematological malignancies: 2.91 %</li> <li>• c.524G&gt;A in haematological malignancies: 216</li> <li>• Total number of haematological malignancies in the database: 7,403</li> </ul>
<b>Solid_Tumour_Freq</b>	Frequency of the variant (cDNA_nomenclature) in solid tumours  e.g.: for variant c.524G>A, the entry will be 4.40 (3,087/70,153) <ul style="list-style-type: none"> <li>• Frequency of c.524G&gt;A in Solid tumours: 4.40 %</li> <li>• c.524G&gt;A in Solid tumours: 3,087</li> <li>• Total number of Solid tumours in the database: 70,153</li> </ul>
<b>Tumour_Freq</b>	Frequency of the variant (cDNA_nomenclature) in tumours only (excluding variants from cell lines, germline and non-neoplastic diseases)  e.g.: for variant c.524G>A, the entry will be 4.33 (3,148/72,829) <ul style="list-style-type: none"> <li>• Frequency of c.524G&gt;A in tumours: 4.33 %</li> <li>• c.524G&gt;A in tumours: 3,148</li> <li>• Total number of tumours in the database: 72,829</li> </ul>
<b>Cell_line_Freq</b>	Frequency of the variant (cDNA_nomenclature) in cell lines only (excluding variants from tumours, germline and non-neoplastic diseases)  e.g.: for variant c.524G>A, the entry will be 3.77 (146/3,864) <ul style="list-style-type: none"> <li>• Frequency of c.524G&gt;A in tumours: 3.77 %</li> <li>• c.524G&gt;A in tumours: 146</li> <li>• Total number of cell lines in the database: 3,864</li> </ul>
<b>Somatic_Freq</b>	Frequency of the variant (cDNA_nomenclature) found as a somatic event  e.g.: for variant c.524G>A, the entry will be 4.13 (3,346/78,639) <ul style="list-style-type: none"> <li>• Frequency of c.524G&gt;A in tumours: 4.26 %</li> <li>• c.524G&gt;A in tumours: 3,346</li> <li>• Total number of cell lines in the database: 78,639</li> </ul>
<b>Germline_Freq</b>	Frequency of the variant (cDNA_nomenclature) found as a germline event  e.g.: for variant c.524G>A, the entry will be 4.47 (52/1,169) <ul style="list-style-type: none"> <li>• Frequency of c.524G&gt;A in tumours: 4.47 %</li> <li>• c.524G&gt;A in tumours: 52</li> <li>• Total number of germline in the database: 1,169</li> </ul> <p>Note of caution 1: for multiple variants, this frequency will be 0, as many of these variants (particularly those associated with carcinogen exposure) are only found as somatic events.</p> <p>Note of caution 2: the germline Brazil mutation p.R337H has been shown to be a founder mutation and has only been included once in the database.</p>

<b>Mutant activities (info)</b>	<p>Data for WAF, MDM2, BAX, 14-3-3-s, AIP, GADD45, NOXA and P53R2 are taken from the publication by Kato et al. (Kato S, Han SY, Liu W, Otsuka K, Shibata H, Kanamaru R, Ishioka C (2003) Understanding the function-structure and function-mutation relationships of p53 tumor suppressor protein by high-resolution missense mutation analysis. Proc Natl Acad Sci U S A 100: 8424-8429).</p> <p>Transactivation was tested using a yeast assay. The residual transcriptional activity of mutant p53 is always compared to wild-type p53 for the same promoter (%).</p> <p><b>Syn:</b> mutation that does not change the amino acid: however, some of these mutations can change splicing or RNA stability.</p> <p><b>Fr:</b> Frameshift mutations. No activity data are available, but it is generally assumed that no p53 is produced.</p> <p><b>Tr:</b> Terminating mutation: No activity data are available, but it is generally assumed that no p53 is produced.</p> <p><b>ND:</b> No data available for this mutant.</p> <p><b>Splice:</b> splice mutation. No activity data are available, but it is generally assumed that no p53 is produced.</p> <p>see <b>Comment frequency</b> for a final assessment</p>
<b>WAF1_Act</b>	Residual transcriptional activity of mutant p53 on the WAF1 promoter (raw data from kato et al.)
<b>MDM2_Act</b>	Residual transcriptional activity of mutant p53 on the MDM2 promoter (raw data from kato et al.)
<b>BAX_Act</b>	Residual transcriptional activity of mutant p53 on the BAX promoter (raw data from kato et al.)
<b>14_3_3_s_Act</b>	Residual transcriptional activity of mutant p53 on the 14-3-3-s promoter (raw data from kato et al.)
<b>AIP_Act</b>	Residual transcriptional activity of mutant p53 on the AIP promoter (raw data from kato et al.)
<b>GADD45_Act</b>	Residual transcriptional activity of mutant p53 on the GADD45 promoter (raw data from kato et al.)
<b>NOXA_Act</b>	Residual transcriptional activity of mutant p53 on the NOXA promoter (raw data from kato et al.)
<b>p53R2_Act</b>	Residual transcriptional activity of mutant p53 on the p52R2 promoter (raw data from kato et al.)
<b>WAF1_percent</b>	Residual transcriptional activity of mutant p53 on the WAF1 promoter (% compared to wild-type p53).
<b>MDM2_percent</b>	Residual transcriptional activity of mutant p53 on the MDM2 promoter (% compared to wild-type p53).
<b>BAX_percent</b>	Residual transcriptional activity of mutant p53 on the BAX promoter (% compared to wild-type p53).
<b>14_3_3_s_percent</b>	Residual transcriptional activity of mutant p53 on the 14-3-3-s promoter (% compared to wild-type p53).
<b>AIP_percent</b>	Residual transcriptional activity of mutant p53 on the AIP promoter (% compared to wild-type p53).
<b>GADD45_percent</b>	Residual transcriptional activity of mutant p53 on the GADD45 promoter (% compared to wild-type p53).
<b>NOXA_percent</b>	Residual transcriptional activity of mutant p53 on the NOXA promoter (% compared to wild-type p53).
<b>p53R2_percent</b>	Residual transcriptional activity of mutant p53 on the p52R2 promoter (% compared to wild-type p53).
<b>Sift_Prediction</b>	Predictive value using Sift
<b>Sift_Score</b>	<p>Predicted functional effect using SIFT algorithm</p> <p><a href="http://sift.jcvi.org/">http://sift.jcvi.org/</a></p> <p>SIFT (Sorting Intolerant From Tolerant) prediction is based on the degree of conservation of amino acid residues in sequence alignments derived from</p>



	<p>closely related sequences.</p> <p>Ranges from 0 to 1. The amino acid substitution is predicted to be damaging when the score is <math>\leq 0.05</math>, and tolerated when the score is <math>&gt; 0.05</math>.</p>
<b>PolyPhen-2 (info)</b>	<p><b>PolyPhen-2</b> is an automatic tool for prediction of the possible impact of an amino acid substitution on the structure and function of a human protein. This prediction is based on a number of features comprising the sequence, phylogenetic and structural information characterizing the substitution.</p> <p><a href="http://genetics.bwh.harvard.edu/pph2/">http://genetics.bwh.harvard.edu/pph2/</a></p> <p><b>Polyphen-2 : prediction_HUMDiv versus prediction_HUMVar</b></p> <p>Two pairs of datasets were used to train and test PolyPhen-2 prediction models. The first pair, <b>HumDiv</b>, was compiled from all damaging alleles with known effects on the molecular function causing human Mendelian diseases, present in the UniProtKB database, together with differences between human proteins and their closely related mammalian homologs, assumed to be non-damaging. The second pair, <b>HumVar</b>, consisted of all human disease-causing mutations from UniProtKB, together with common human nsSNPs (MAF<math>&gt;1\%</math>) without annotated involvement in disease, which were treated as non-damaging.</p> <p>The user can choose between HumDiv- and HumVar-trained PolyPhen-2 models. Diagnostics of Mendelian diseases requires distinguishing mutations with drastic effects from all the remaining human variation, including abundant mildly deleterious alleles. Thus, HumVar-trained model should be used for this task. In contrast, HumDiv-trained model should be used for evaluating rare alleles at loci potentially involved in complex phenotypes, dense mapping of regions identified by genome-wide association studies, and analysis of natural selection from sequence data, where even mildly deleterious alleles must be treated as damaging.</p> <p>More info on the polyphen web site : <a href="http://genetics.bwh.harvard.edu/pph2/dokuwiki/overview">http://genetics.bwh.harvard.edu/pph2/dokuwiki/overview</a></p>
<b>Polyphen-2_HumVar</b>	See above for more info
<b>Polyphen-2_HumDiv</b>	See above for more info; this prediction is less accurate than HumVar for TP53
<b>Mutassessor_prediction:</b>	Functional impact of a variant : predicted functional (high, medium), predicted non-functional (low, neutral).
<b>Mutassessor_score:</b>	<p>Predicted functional effect using Mutassessor algorithm</p> <p><a href="http://mutationassessor.org/">http://mutationassessor.org/</a></p> <p>B. Reva, Y. Antipin, C. Sander, <i>Nucleic Acids Res</i> <b>39</b>, e118 (2011).</p> <p>Functional impact combined score</p> <p>The <b>default score cutoff</b> is currently set at -1.938 for classification (i.e. High or medium vs low or neutral).</p>
<b>Provean_prediction</b>	Prediction - deleterious or neutral (using default cutoff at -2.5)
<b>Provean_Score</b>	<p>Predicted functional effect using Provean algorithm</p> <p><a href="http://provean.jcvi.org/index.php">http://provean.jcvi.org/index.php</a></p> <p><b>PROVEAN (Protein Variation Effect Analyzer)</b> is a software tool which predicts whether an amino acid substitution or indel has an impact on the biological function of a protein.</p> <p>PROVEAN introduces a <b>delta alignment score</b> based on the reference and variant versions of a protein query sequence with respect to sequence homologues collected from the NCBI NR protein database through BLAST.</p>

	<p>T</p> <p>For maximum separation of deleterious and neutral variants for all 4 classes of human protein variants, the <b>default score cutoff</b> is currently set at -2.5 for binary classification (i.e. deleterious vs neutral).</p>
Condel	<p>Predicted functional effect using Condel algorithm</p> <p><a href="http://bg.upf.edu/fannsdb/">http://bg.upf.edu/fannsdb/</a></p>
Condel_Score	<p>Predicted functional effect using Condel algorithm</p> <p><a href="http://bg.upf.edu/fannsdb/">http://bg.upf.edu/fannsdb/</a></p>
MutPred_Splice_General_Score	<p><a href="http://mutdb.org/mutpredsplice/about.htm">http://mutdb.org/mutpredsplice/about.htm</a></p> <p>The MutPred Splice outputs are:</p> <ol style="list-style-type: none"> <li>1, <b>General Score</b>, which is the probability that the variant disrupts splicing. We use a general score <math>\geq 0.6</math> to identify a variant which disrupts splicing. e.g. general score <math>\geq 0.6</math> labels a variant as a Splice Affecting Variant (SAV) e.g. general score <math>&lt; 0.6</math> labels a variant as a Splice Neutral Variant (SNV)</li> <li>2, Additional supporting evidence is provided by a <b>confident hypothesis</b> about the splicing mechanism disrupted.</li> </ol> <p>Practical advice</p> <p>MutPred Splice can be used to prioritise your dataset into three partitions:</p> <ol style="list-style-type: none"> <li>1, High Confident calls of splicing variants - predicted SAV (general score <math>\geq 0.6</math>) where a confident hypothesis is available.</li> <li>2, Confident calls of splicing variants - predicted SAV (general score <math>\geq 0.6</math>) where a confident hypothesis not available.</li> <li>3, Not predicted to disrupt splicing (SNV) (general score <math>&lt; 0.6</math>).</li> </ol>
MutPred_Splice_Prediction_Label	<p>See above and <a href="http://mutdb.org/mutpredsplice/about.htm">http://mutdb.org/mutpredsplice/about.htm</a></p>
MutPred_Splice_Confident_Hypotheses	<p>See above and <a href="http://mutdb.org/mutpredsplice/about.htm">http://mutdb.org/mutpredsplice/about.htm</a></p>
SIFT_converted_rankscore	<p>SIFTori scores were first converted to <math>SIFT_{new} = 1 - SIFT_{ori}</math>, then ranked among all <math>SIFT_{new}</math> scores in dbNSFP. The rankscore is the ratio of the rank the <math>SIFT_{new}</math> score over the total number of <math>SIFT_{new}</math> scores in dbNSFP. If there are multiple scores, only the most damaging (largest) rankscore is presented. The rankscores range from 0.02654 to 0.87932.</p>
Polyphen2_HDIV_rankscore	<p>Polyphen2 HDIV scores were first ranked among all HDIV scores in dbNSFP. The rankscore is the ratio of the rank the score over the total number of the scores in dbNSFP. If there are multiple scores, only the most damaging (largest) rankscore is presented. The scores range from 0.02656 to 0.89917</p>
Polyphen2_HVAR_rankscore	<p>Polyphen2 HVAR scores were first ranked among all HVAR scores in dbNSFP. The rankscore is the ratio of the rank the score over the total number of the scores in dbNSFP. If there are multiple scores, only the most damaging (largest) rankscore is presented. The scores range from 0.01281 to 0.9711</p>
LRT_converted_rankscore	<p>LRTori scores were first converted as <math>LRT_{new} = 1 - LRT_{ori} \cdot 0.5</math> if <math>\Omega &lt; 1</math>, or <math>LRT_{new} = LRT_{ori} \cdot 0.5</math> if <math>\Omega \geq 1</math>. Then <math>LRT_{new}</math> scores were ranked among all <math>LRT_{new}</math> scores in dbNSFP. The rankscore is the ratio of the rank over the total number of the scores in dbNSFP. The scores range from 0.00166 to 0.85682.</p>
MutationTaster_converted_rankscore	<p>The MTori scores were first converted: if the prediction is "A" or "D" <math>MT_{new} = MT_{ori}</math>; if the prediction is "N" or "P", <math>MT_{new} = 1 - MT_{ori}</math>. Then <math>MT_{new}</math> scores were ranked among all <math>MT_{new}</math> scores in dbNSFP. If there are multiple scores of a SNV, only the largest <math>MT_{new}</math> was used in ranking. The rankscore is the ratio of the rank of the score over the total number of <math>MT_{new}</math> scores in dbNSFP. The scores range from 0.08979 to 0.81033.</p>
MutationAssessor_rankscore	<p>MAori scores were ranked among all MAori scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of MAori scores in dbNSFP. The scores range from 0 to 1.</p>
FATHMM_rankscore	<p>FATHMMori scores were first converted to <math>FATHMM_{new} = 1 - (FATHMM_{ori} + 16.13) / 26.77</math>, then ranked among all <math>FATHMM_{new}</math> scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of <math>FATHMM_{new}</math> scores in dbNSFP. If there are multiple scores,</p>

	only the most damaging (largest) rankscore is presented. The scores range from 0 to 1.
<b>MetaSVM_rankscore</b>	MetaSVM scores were ranked among all MetaSVM scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of MetaSVM scores in dbNSFP. The scores range from 0 to 1.
<b>MetaLR_rankscore</b>	MetaLR scores were ranked among all MetaLR scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of MetaLR scores in dbNSFP. The scores range from 0 to 1.
<b>VEST3_rankscore</b>	VEST3_rankscore: VEST3 scores were ranked among all VEST3 scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of VEST3 scores in dbNSFP. In case there are multiple scores for the same variant, the largest score (most damaging) is presented. The scores range from 0 to 1. Please note VEST score is free for non-commercial use. For more details please refer to <a href="http://wiki.chasmssoftware.org/index.php/SoftwareLicense">http://wiki.chasmssoftware.org/index.php/SoftwareLicense</a> . Commercial users should contact the Johns Hopkins Technology Transfer office.
<b>PROVEAN_converted_rankscore</b>	PROVEAN <sub>Nori</sub> were first converted to PROVEAN <sub>new</sub> =1-(PROVEAN <sub>Nori</sub> +14)/28, then ranked among all PROVEAN <sub>new</sub> scores in dbNSFP. The rankscore is the ratio of the rank of the PROVEAN <sub>new</sub> score over the total number of PROVEAN <sub>new</sub> scores in dbNSFP. If there are multiple scores, only the most damaging (largest) rankscore is presented.
<b>phyloP46way_primate_rankscore</b>	phyloP46way_primate scores were ranked among all phyloP46way_primate scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phyloP46way_primate scores in dbNSFP.
<b>phyloP46way_placental_rankscore</b>	phyloP46way_placental scores were ranked among all phyloP46way_placental scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phyloP46way_placental scores in dbNSFP.
<b>GERP_RS_rankscore</b>	GERP++ RS scores were ranked among all GERP++ RS scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of GERP++ RS scores in dbNSFP.
<b>SiPhy_29way_logOdds_rankscore</b>	SiPhy_29way_logOdds_rankscore: SiPhy_29way_logOdds scores were ranked among all SiPhy_29way_logOdds scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of SiPhy_29way_logOdds scores in dbNSFP
<b>Evol_Score_II</b>	Evolutionary Acation score score as defined by Lichtarge et al. <a href="https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4383697">https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4383697</a> <a href="http://mammoth.bcm.tmc.edu/ETserver.html">http://mammoth.bcm.tmc.edu/ETserver.html</a>
<b>RFscore_II</b>	MutPred is based upon SIFT and a gain/loss of 14 different structural and functional properties. For instance, gain of helical propensity or loss of a phosphorylation site. It was trained using the deleterious mutations from the Human Gene Mutation Database and neutral polymorphisms from Swiss-Prot]. Current version of MutPred is 1.2. The update consists of replacing SIFT score by a more stable version of code that calculates evolutionary conservation  <a href="http://mutpred.mutdb.org/about.html">http://mutpred.mutdb.org/about.html</a>
<b>phyloP46way_primate_rankscore</b>	phyloP46way_primate scores were ranked among all phyloP46way_primate scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phyloP46way_primate scores in dbNSFP
<b>phyloP46way_placental_rankscore</b>	phyloP46way_placental scores were ranked among all phyloP46way_placental scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phyloP46way_placental scores in dbNSFP
<b>phyloP100way Vertebrate_rankscore</b>	phyloP100way Vertebrate scores were ranked among all phyloP100way Vertebrate scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phyloP100way Vertebrate scores in dbNSFP.
<b>phastCons46way_primate_rankscore</b>	phastCons46way_primate scores were ranked among all phastCons46way_primate scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phastCons46way_primate scores in dbNSFP

<b>phastCons46way_placental_rankscore</b>	phastCons46way_placental scores were ranked among all phastCons46way_placental scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phastCons46way_placental scores in dbNSFP
<b>phastCons100way_vertebrate_rankscore</b>	phastCons100way_vertebrate_rankscore: phastCons100way_vertebrate scores were ranked among all phastCons100way_vertebrate scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of phastCons100way_vertebrate scores in dbNSFP.
<b>GERP_RS_rankscore</b>	GERP++ RS scores were ranked among all GERP++ RS scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of GERP++ RS scores in dbNSFP.
<b>CADD_raw_rankscore</b>	CADD raw scores were ranked among all CADD raw scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of CADD raw scores in dbNSFP. Please note the following copyright statement for CADD: "CADD scores ( <a href="http://cadd.gs.washington.edu/">http://cadd.gs.washington.edu/</a> ) are Copyright 2013 University of Washington and Hudson-Alpha Institute for Biotechnology (all rights reserved) but are freely available for all academic, non-commercial applications. For commercial licensing information contact Jennifer McCullar ( <a href="mailto:mccullaj@uw.edu">mccullaj@uw.edu</a> ).
<b>DANN_rank_score</b>	DANN scores were ranked among all DANN scores in dbNSFP. The rankscore is the ratio of the rank of the score over the total number of DANN scores in dbNSFP.
<b>Mutalyzer_comment</b>	Comment generated by mutalyzer when the mutation has not been correctly annotated.
<b>CHROM</b>	User data
<b>POS</b>	User data
<b>ID</b>	User data
<b>REF</b>	User data
<b>ALT</b>	User data